#### **Case Report**

# Vitamin D Toxicity; Stored and Released from Adipose Tissue?

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#### Abstract

Vitamin D is a pro-hormone that plays an important role in body calcium regulation. Vitamin D toxicity occurs rarely due to its wide therapeutic index. A 38-year-old Iranian man was admitted to our hospital with a diagnosis of vitamin D toxicity. Laboratory test revealed hypercalcemia and elevated 25-hydroxyvitamin D (25(OH)D) level. Despite the cessation of vitamin D intake and diuretic treatment, he presented four months later with high 25(OH)D level and similar clinical features. Due to the potential release of vitamin D from adipose tissue, serial monitoring of 25(OH)D level is recommended.

Keywords: Vitamin D, toxicity, adipose tissue

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# Introduction

V itamin D is a fat-soluble vitamin with anti-inflammatory and immune-modulating properties which plays an important role in calcium homeostasis and bone mineral metabolism.<sup>1,2</sup> Vitamin D has two distinct forms; Ergocalciferol or vitamin D<sub>2</sub> and vitamin D<sub>3</sub> or cholecalciferol. Vitamin D<sub>2</sub> is present in the diet (some plants and fish) and vitamin D<sub>3</sub> is synthesized from 7-dehydrocholesterol in the skin. Vitamin D, from diet and skin, is then transported to liver where it undergoes 25-hydroxylation, yielding 25-hydroxyvitamin D [25(OH)D].<sup>3–5</sup> which is widely distributed in the body and is the major circulating metabolite of vitamin D.<sup>4.5</sup>

The biologically active form of vitamin D is synthesized in the kidney by conversion of 25(OH)D to 1,25-dihydroxyvitamin D  $[1,25(OH)_2D]$ .  $1,25(OH)_2D$  promotes intestinal calcium absorption and maintains adequate serum calcium and phosphate concentrations to enable normal mineralization.<sup>3-5</sup> However, if there are enough supplies of calcium and phosphorus in the body, 25(OH)D is converted to 24,25-dihydroxyvitamin D  $[24,25(OH)_2D]$  whose role is still unknown but is believed to be a potential biomarker of vitamin D catabolism, an intermediate in the inactivation-excretion process.<sup>6</sup>

Adipose tissue is considered as the major repository of vitamin D in the body; nevertheless, it is clearly distributed in other tissues like muscles.<sup>6,7</sup> The serum concentration of 25(OH)D is an appropriate indicator of vitamin D status, reflecting both the amount of vitamin D absorbed from diet and produced cutaneously.<sup>8</sup> This is due to its long serum half life (15 days) and high circulatory levels compared to 1,25(OH)<sub>2</sub>D (half life of 4-6 hours).<sup>3,8</sup> However, 25(OH)D is not an indicator of stored vitamin D.<sup>8</sup>

The major source of vitamin D for most humans is exposure to sunlight. Few foods naturally contain vitamin D (oily fish such as salmon, mackerel, etc), and nowadays vitamin D deficiency is recognized as a pandemic.9 Vitamin D deficiency will cause rickets in children and is associated with increased risk of cancer, autoimmune disease, hypertension and osteoporosis.<sup>10</sup> The daily recommended requirement for vitamin D intake is 200 IU/day for children, 400 IU/day for adults below 50 years of age, 600 IU/day between ages 50 and 70 and thereafter 800 IU/D.1,4 The tolerable upper intake level of vitamin D likely to cause no potential harm (UL) is currently defined as 2000 IU/day both in North America and Europe.<sup>11</sup> Because of the wide therapeutic index, vitamin D toxicity occurs rarely but there are still iatrogenic cases due to self-administered medication or, to a lesser extent, fortified food products.<sup>10,12</sup> Hypercalcemia is mainly responsible for vitamin D toxicity symptoms. The symptoms have a wide range including headache, bone pains, arrhythmia, loss of appetite, frequent urination, nervousness, itching, kidney stones and gastrointestinal symptoms such as diarrhea, nausea, vomiting and constipation.13,14

In this paper, we present a classic case of vitamin D toxicity whose 25(OH)D level was elevated once again after four months despite the fact that supplements and vitamin D intake were ceased and the patient underwent treatment.

## **Case Report**

A 38-year-old Iranian male was admitted to our hospital because of bloodshot eyes, arthralgia, ostealgia, and weight loss for 18 months. The patient was first presented to his clinician with a complaint of non-itchy red eye without discharge or any other effect on his vision. He had received treatment for conjunctivitis several times. Two months later, the patient experienced generalized pruritus without any other dermatological symptoms, unresponsive to common medication. After three months, he complained of a progressive ostealgia, starting at lower extremities, particularly in the plantar regions, and fatigue, weakness, arthralgia and hypohidrosis. He also mentioned a significant weight loss, 16.5 kilograms over the past 18 months.

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No sign of arthritis was observed. He also mentioned a mild hearing loss for a couple of months prior to his admission.

The patient is a professional bodybuilder since early adolescence. He has consumed various types of minerals, vitamins and protein supplements. Remarkably, he gave a history of weekly intramuscular (IM) injections of vitamin D (300,000 IU) for 10 years, which subsequently increased to two IM injections per week as his symptoms aggravated.

On physical examination, he was conscious and well-oriented. His vital signs were normal as follows: blood pressure 132/78 mmHg, pulse rate 72 per minute, respiratory rate 16 per minute and temperature 36.8°C. His conjunctiva was slightly pale. Heart, chest and abdomen were found normal. Extremities were also normal on neurological examination with no sign of edema or clubbing.

On ECG, he had short QT intervals (Figure 1). In ophthalmology assessment, keratopathy band was observed (Figure 2). Myringosclerosis, conductive hearing loss and type E tympanometry were reported in his ENT consultation. The CT scan of temporal bone showed no pathology. During his stay, due to a right flank pain, an abdominopelvic spiral CT scan without contrast was performed and several nephrolithiases and nephrocalcinoses were found (Figure 3). He was treated

with transurethral ureterolithotripsy (TUL) because of further obstructive symptoms.

On laboratory investigation, hypercalcemia and high levels of BUN and creatinine were reported (Table 1). On further assessment, the 25(OH)D level was reported at 310 ng/mL (normal range: 20–100 ng/mL). Therefore, a diagnosis of vitamin D toxicity induced acute kidney injury and hypercalcemia was stated for him. The intake of all supplements and vitamin D was stopped and he was treated with intravenous fluid (2/3 of his urine output) and furosemide 40 mg three times per day. After 25 days, his creatinine and calcium levels gradually declined and his symptoms improved. The 25(OH)D level was reported at 105 ng/mL and he was discharged. He was advised not to take any supplements or vitamin D injections afterwards.

Despite discontinuing vitamin D intake, the patient presented four months later with severe ostealgia, 25(OH)D level of 500 ng/mL, and high creatinine and calcium levels (Table 1). He was admitted once again and further work up was done. Hepatitis B surface antigen, hepatitis C antibody, and human immunodeficiency virus antibody were negative. Skull X-ray was normal. Urine and blood protein electrophoresis were normal without spike. A thorough panel of the rheumatologic tests was performed with no abnormality found (Table 2).



Figure 1. Electrocardiogram. Short QT intervals.



Figure 2. Keratopathy band.



Figure 3. Abdominopelvic spiral CT scan, several nephrolithiases

| Table 1. Laboratory | y test result. |
|---------------------|----------------|
|---------------------|----------------|

| Variable                               | Normal range | First admission | Second admission | Discharge |
|--|--------------|-----------------|------------------|-----------|
| WBC (per man <sup>3</sup> )            | 4000-10,000  | 7000            | 9500             | 9200      |
| Hb (g/dL)                              | 14–18        | 10.4            | 10.8             | 10.9      |
| Plt (× $10^{3}$ per mar <sup>3</sup> ) | 140–440      | 275             | 201              | 198       |
| MCV (fL)                               | 77–97        | 82.4            | 86.6             | 87.2      |
| MCH (pg/cell)                          | 26–32        | 27.5            | 28.4             | 28.6      |
| BUN (mg/dL)                            | 5–23         | 28              | 51               | 31        |
| Creatinine(mg/dL)                      | 0.5-1        | 4.3             | 3.7              | 2.6       |
| Serum Albumin(g/dL)                    | 3.5–5        |                 | 3.5              | 3.7       |
| Ca (mg/dL)                             | 8.6-10.2     | 13.8            | 14.1             | 10.2      |
| P (mg/dL)                              | 2.5–5        | 4.1             | 5.1              | 4.2       |
| Na (mEq/L)                             | 136–145      | 136             | 137              | 142       |
| K (mEq/L)                              | 3.7–5.5      | 3.9             | 4.4              | 4.5       |
| Mg (mg/dL)                             | 1.5-2.5      | 2.4             | 2.2              | 1.8       |
| Serum Iron (mcg/dL)                    | 60–170       | 70              | 42               |           |
| TIBC (mcg/dL)                          | 240-450      | 375             | 385              |           |
| Ferritin (ng/mL)                       | 17–390       | 801             | 548              |           |
| UA-SG                                  |              | 1015            | 1007             |           |
| UA WBC                                 |              | 35–40           | 13–15            |           |
| UA RBC                                 |              | 3–4             | 4–6              |           |
|  |              |                 |                  |           |

WBC = white blood cell count, Hb = hemoglobin, Plt = platelet count, MCV = mean corpuscular volume, MCH = mean corpuscular hemoglobin, BUN = blood urea nitrogen, Ca = calcium, P = phosphorous, Na = sodium, K = potassium, Mg = magnesium, TIBC = total iron binding capacity, UA-SG = urine analysis specific gravity, UA RBC = urine analysis red blood cell, UA WBC = urine analysis white blood cell.

#### Table2. Rheumatologic panel test.

| Variable  | Normal range | Patient |  |  |
|---|--------------|---------|--|--|
| T4  | 4-13         | 11.7    |  |  |
| Т3  | 80–250       | 108     |  |  |
| T3RU  | 30–40        | 32      |  |  |
| TSH   | 0.3–5        | 1.1     |  |  |
| 1,84-PTH  | 6.5–37       | <4      |  |  |
| СРК   | 20–200       | 94      |  |  |
| Aldolase  | <8.8         | 3.4     |  |  |
| ANA   | <1=Neg       | Neg     |  |  |
| Anti-ds DNA   | <1=Neg       | Neg     |  |  |
| SSA/Ro IgG  | <1=Neg       | Neg     |  |  |
| SSB/La IgG  | <1=Neg       | Neg     |  |  |
| Anti-Mpo  | <1=Neg       | Neg     |  |  |
| Anti-PR3  | <1=Neg       | Neg     |  |  |
| Anti-ccp  | <1=Neg       | Neg     |  |  |
| ESR   | 0–22         | 38      |  |  |
| CRP   |              | Neg     |  |  |
| RF  |              | Neg     |  |  |
| $\alpha_1$ Globulin   | 2–6          | 5.5     |  |  |
| a, Globulin   | 8–13         | 12.6    |  |  |
| γ Globulin  | 13–19        | 16      |  |  |
| Serum Albumin   | 3.5–5.5      | 3.5     |  |  |
| Alb/Glb   | 1.2–2.4      | 0.95    |  |  |
| T3RU = T3 resin uptake. TSH = thyroid stimulating hormone. PTH = parathyroid hormone. CPK = creatinine phosphokinase. ANA = anti- nuclear antibody. |              |         |  |  |

 $13 \text{ RO} = 13 \text{ resin uptake, } 1SH = thyroid stimulating hormone, } PTH = parathyroid hormone, CPK = creatinine phosphokinase, ANA = anti-nuclear antibody, Anti-ds DNA = anti double stranded deoxyribonucleic acid antibody, SSA/Ro = anti-Sjögren's syndrome A antibody, SSB/La = anti-Sjögren's syndrome B antibody, Anti-Mpo = anti-myeloperoxidase antibody, Anti-PR3 = anti-proteinase-3 antibody; Anti-ccp = anti-cyclic citrullinated peptide antibody, ESR = erythrocyte sedimentation rate, CRP = C-reactive protein, RF = rheumatoid factor, Alb/Glb = albumin to Globulin ration.$ 

His second hospital stay lasted 27 days and he was treated with intravenous fluid (2/3 of his urine output) and furosemide 40 m three times per day. The 25(OH)D level declined to 263 ng/mL and the calcium level to 10.2 mg/dL. He was discharged with a prescription of calcium-restricted diet, and furosemide 40 mg three times per day. Further follow up was recommended.

#### Discussion

Vitamin D is a principal pro-hormone in body which is affected by many factors including dietary intake, supplement, latitude, skin color and sun exposure.<sup>15</sup> Vitamin D toxicity rarely happens due to the wide therapeutic index, tightly regulated synthesis of 1,25(OH)<sub>2</sub>D and the characteristics of adipose tissue as a depot for storage and release of vitamin D.<sup>2,3</sup> In this case report, a patient with classic symptoms of hypervitaminosis D is presented whose 25(OH)D level was reduced after cessation of all supplements and proper treatment; however, after four months, the 25(OH)D level was remarkably elevated and the patient's symptoms returned.

Vitamin D, regardless of origin, is an inactive pro-hormone. It is transported to the liver by vitamin D binding protein (DBP) and it is metabolized to 25(OH)D by a cytochrome P450 (CYP) enzyme, likely CYP2R1.<sup>6,16</sup> The further hydroxylation reaction occurs in the kidney and is tightly regulated by the induction of CYP27B1 enzyme.<sup>16</sup> The production of CYP27B1 is stimulated by parathyroid hormone (PTH) which is secreted in response to hypocalcemia. CYP27B1 is suppressed by calcitriol  $(1,25(QH)_2$  D) and hyperphosphatemia.<sup>16</sup> This metabolic step constitutes the basis of vitamin D regulation that is central to calcium and phosphate homeostasis.

Vitamin D is a fat soluble vitamin and it is primarily excreted in bile; however, the exact mechanism or the related excretion products are not clear.<sup>17</sup> Adipose tissue is the predominant storage site for vitamin D. Following vitamin D intake, it is rapidly distributed in adipose tissue and then slowly released into systemic circulation. These characteristics of accumulation and release could have the potential as a protective mechanism against toxicity, maintaining the balance of plasma concentration of vitamin D level.<sup>18,19</sup> The issue of storage could be confirmed by the fact that inducing a deficiency state in a laboratory animal takes time.<sup>20</sup> Furthermore, the literature states an estimation of four months for biological activity of vitamin D in human plasma after cessation of large doses.<sup>21</sup>

Adiposity is inversely associated with 25 (OH)D level and obese people have lower 25(OH)D level which might be due to decreased bioavailability of vitamin D after it is deposited in fat tissue.19 Vitamin D is a fat soluble vitamin stored and sequestrated in adipose tissue.<sup>22</sup> Conversely, Blum et al.<sup>22</sup> found a positive correlation between serum and fat tissue D3 levels using liquid chromatography mass spectrometry (LC/MS). Heaney et al.23 indicated that the amount of vitamin D in adipose tissue is too small to affect 25(OH)D level during weight loss. In another study conducted by Pramyothin et al.,24 despite remarkable loss of fat mass after Roux-en-Y gastric bypass surgery, vitamin D in adipose tissue did not contribute significantly to serum 25(OH) D level. Conversely, Lin et al.<sup>25</sup> observed a positive association between adipose tissue and systemic 25(OH)D concentration, suggesting adipose tissue as a storage site that releases vitamin D during weight loss. Therefore, further investigation is warranted to ascertain whether vitamin D is stored in body and then released or it is sequestered in adipose tissue in obese individuals.

Vitamin D supplements are available over the counter and nowadays, there is a growing concern regarding "vitamin D deficiency" and the consequent potential issues, as in this case report, the patient consumed vitamin D supplements for approximately 10 years without any medical problem. Thus, careful dosing of vitamin D is important. Furthermore, considering the role of adipose tissue as the main storage site that releases vitamin D, serial monitoring of 25(OH)D level could be a good indicator of vitamin D status in patients treated for vitamin D toxicity.

## References

- 1. Heaney RP. Vitamin D in health and disease. *Clin J Am Soc Nephrol.* 2008; 3(5): 1535 1541.
- 2. Alshahrani F, Aljohani N. Vitamin D: deficiency, sufficiency and tox-

icity. Nutrients. 2013; 5(9): 3605-3616.

- Jones G. Pharmacokinetics of vitamin D toxicity. Am J Clin Nutr. 2008; 88(2): 582S – 586S.
- Bansal RK, Tyagi P, Sharma P, Singla V, Arora V, Bansal N, et al. Iatrogenic hypervitaminosis D as an unusual cause of persistent vomiting: a case report. *J Med Case Rep.* 2014; 26(8): 74.
- 5. Heaney RP. Vitamin D: criteria for safety and efficacy. *Nutr Rev.* 2008; 66(10 Suppl 2): S178 181.
- Bosworth CR, Levin G, Robinson-Cohen C, Hoofnagle AN, Ruzinski J, Young B, et al. The serum 24,25-dihydroxyvitamin D concentration, a marker of vitamin D catabolism, is reduced in chronic kidney disease. *Kidney Int.* 2012; 82(6): 693 – 700.
- Mutt SJ, Hyppönen E, Saarnio J, Järvelin MR, Herzig KH. Vitamin D and adipose tissue-more than storage. *Front Physiol*. 2014; (5): 228.
- van den Ouweland JM, Beijers AM, Demacker PN, van Daal H. Measurement of 25-OH-vitamin D in human serum using liquid chromatography tandem-mass spectrometry with comparison to radioimmunoassay and automated immunoassay. *J Chromatogr B Analyt Technol Biomed Life Sci.* 2010; 878(15-16): 1163 – 1168.
- Holick MF, Chen TC. Vitamin D deficiency: a worldwide problem with health consequences. *Am J Clin Nutr.* 2008; 87(4): 1080S – 1086S.
- Kulie T, Groff A, Redmer J, Hounshell J, Schrager S. Vitamin D: an evidence-based review. J Am Board Fam Med. 2009; 22(6): 698 – 706.
- Vieth R. Vitamin D toxicity, policy, and science. J Bone Miner Res. 2007; (22 Suppl 2): V64 – V68.
- Koul PA, Ahmad SH, Ahmad F, Jan RA, Shah SU, Khan UH. Vitamin d toxicity in adults: a case series from an area with endemic hypovitaminosis d. *Oman Med J.* 2011; 26(3): 201 – 204.
- Schwalfenberg G. Not enough vitamin D: health consequences for Canadians. *Can Fam Physician*. 2007; 53(5): 841 – 854.
- Genzen JR. Hypercalcemic crisis due to vitamin D toxicity. *Lab Med.* 2014; 45(2): 147 – 150.
- Holick MF. Vitamin D: a D-Lightful health perspective. Nutr Rev. 2008; 66(10 Suppl 2): S182 – S194.
- Omdahl JL, Morris HA, May BK. Hydroxylase enzymes of the vitamin D pathway: expression, function, and regulation. *Annu Rev Nutr.* 2002; 22: 139 – 166.
- DeLuca HF. Metabolism of vitamin D: current status. *Am J Clin Nutr.* 1976; 29(11): 1258 – 1270.
- Rosenstreich SJ, Rich C, Volwiler W. Deposition in and release of vitamin D3 from body fat: evidence for a storage site in the rat. *J Clin Invest.* 1971; 50(3): 679 – 687.
- Liel Y, Ulmer E, Shary J, Hollis BW, Bell NH. Low circulating vitamin D in obesity. *Calcif Tiss Int.* 1988; (43): 199 – 201.
- Johnson DW, Palmer LS. Individual and breed variations in pigs on rations devoid of vitamin D. J Agr Res. 1939; (58): 929 – 940.
- Warkany J, Guest GM, Grabill FJ. Estimation of vitamin D in blood serum. Vitamin D in human serum during and after periods of ingestion of large doses of vitamin D. *J Lab Clin Med.* 1942; (27): 557.
- 22. Blum M, Dolnikowski G, Seyoum E, Harris SS, Booth SL, Peterson J, et al. Vitamin D(3) in fat tissue. *Endocrine*. 2008; 33(1): 90 94.
- Heaney RP, Horst RL, Cullen DM, Armas LA. Vitamin D3 distribution and status in the body. *J Am Coll Nutr*. 2009; 28(3): 252 – 256.
- Pramyothin P, Biancuzzo RM, Lu Z, Hess DT, Apovian CM, Holick MF. Vitamin D in adipose tissue and serum 25-hydroxyvitamin D after roux-en-Y gastric bypass. *Obesity (Silver Spring)*. 2011; 19(11): 2228 – 2234.
- Lin E, Armstrong-Moore D, Liang Z, Sweeney JF, Torres WE, Ziegler TR, et al. Contribution of adipose tissue to plasma 25-hydroxyvitamin D concentrations during weight loss followinggastric bypass surgery. *Obesity (Silver Spring)*. 2011; 19(3): 588 – 594.